

ANALYSIS OF POLYMORPHISM OF BETA CASEIN OF SLOVAK PINZGAU CATTLE BY PCR-RFLP FOR ALLELS A1 AND A2

ANALIZA POLIMORFISMULUI GENEI β -CAZEINEI PRIN METODA PCR-RFLP PENTRU ALELELE A1 ȘI A2 LA TAURINELE DE RASĂ PINZGAU SLOVACĂ

MARTINA MILUCHOVÁ, ANNA TRAKOVICKÁ, M. GÁBOR

Slovak University of Agriculture in Nitra, martina.miluchova@centrum.sk,

The work was oriented to identification of β -casein gene polymorphism and analysis of genotype structure in population of Slovak Pinzgau cattle. The material involved 89 cattle. Bovine genomic DNA was isolated by fenol-chlorophorm deprotenization and ethanol precipitation and used in order to estimate β -casein genotypes by means of PCR-RFLP method. The PCR products were digested with DdeI restriction enzyme. In the population included in the study there were homozygote genotype A1A1 (27 animals), heterozygote genotype A1A2 (46 animals) and homozygote genotype A2A2 (16 animals). In the total population of cattle heterozygotes A1A2 – 0.5168 were the most frequent, while homozygotes A2A2 – 0.1798 were the least frequent ones. This suggests a slight superiority of allele A1 – 0.5618.

Keywords: cattle, PCR-RFLP, β -casein

Introduction

The beta - casein (CSN2) constitutes up to 45 % of the casein of bovine milk. CSN2 is localised in bovine chromosome 6 (Ferretti, Leone, Sgaramella, 1990). The primary sequence of β - casein gene reported Ribadeau-Dumas et al. (1972). Single - polypeptide chain of this protein containing 209 residues with molecular weight of 23983.

The β - casein gene has 12 known genetic variants in the coding sequence of the gene: A, B, C (Aschaffenburg, 1961), A1, A2, A3 (Kiddy, 1966), D (Aschaffenburg, 1968), E (Voglino, 1972), F (Visser, 1991), X (Visser et al., 1995), G (Chin,Ng-Kway-Hang, 1997), H (Han et al., 2000), I (Jann et al., 2002).

The most common forms of beta-casein in dairy cattle breeds are A1 and A2, while B is less common, and A3 and C are rare (Farrell et al., 2004; Kamiński, Cieślińska, Kostyra, 2007; Keating et al., 2008). The β - casein A1 and B variants differ from the A2 variant at position 67 where a histidine replaces a proline. In addition, the B variant differs from the A1 variant in a substitution of argine for serine at position 122. Importantly, it is the change to histidine at position 67 that

has the potential to result in cleavage occurring upon digestion and a bioactive peptide, beta-casomorphin potentially being liberated (Stewart et al., 1987; Damiani et al., 1992; Lien et al., 1992).

The β -casein A1 variant was associated with the incidence of diabetes mellitus 1st type, coronary heart disease and autism (Elliot et al., 1988). Fat percentage was estimated with CSN2 genotype A1A1. CSN2 genotype significantly influenced milk yield, fat yield, and protein yield with the highest yields obtained for the genotype A2A2. The A2 variant reduces serum cholesterol. The A3 variant of bovine CSN2 is associated with higher milk yield while higher protein, casein yields and fat are associated with the B variant. (Panicke, Freyer and Erhardt, 1997).

Materials and Methods

The material involved 89 cattle. Bovine genomic DNA was isolated by phenol-chloroform deproteinization and ethanol precipitation (Sambrook et al., 1989) and used in order to estimate CSN2 genotypes by means of PCR-RFLP method.

DNA primers described by McLachlan (2006) were used to PCR amplification: forward primer 5'- CCT TCT TTC CAG GAT GAA CTC CAG G-3' and reverse primer 5' - GAG TAA GAG GAG GGA TGT TTT GTG GGA GGC TCT- 3'.

The PCR reaction elaborated by McLachlan (2006) was modified.

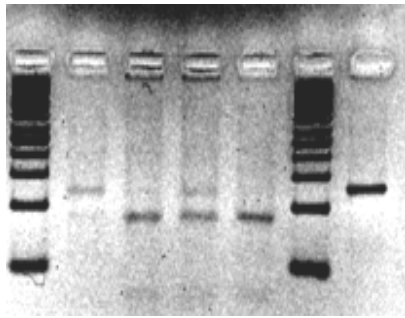
The reaction mixture in the total volume 25 μ l containing 50 ng DNA, 1.5 U Taq polymerase (Fermentas), 3 mM MgCl₂, 200 μ M dNTP, 5 pM of each primer, 0,006 mg BSA. The following amplification parameters were applied: 95°C for 5 minutes followed by 30 cycles: 95 °C for 40 seconds, 58 °C for 60 seconds, 72 °C for 90 seconds. The reaction was completed by the final synthesis: 72 °C for 10 minutes.

The PCR products of 121 bp were digested with 5 units of the *DdeI* restriction enzyme (Promega). Restriction digestion fragments were loaded on 3 % agarose gel (Invitrogen) containing ethidium bromide and the gel were analyzed in the UV rays.

Results and Discussion

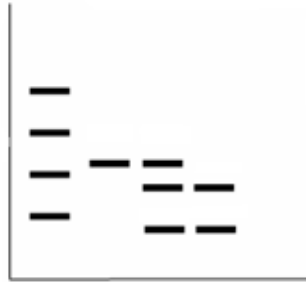
DdeI digestion of the PCR product was analyzed for 3% agarose-gel electrophoresis. Allele A1 produced 121 bp fragments, and allele A2 produced a 86 bp and 35 bp fragments as the PCR-RFLP.

In the population of Slovak Pinzgau cattle we detected all genotypes: homozygote genotype A1A1 (121 bp) 27 animals, heterozygote genotype A1A2 (121 bp, 86 bp, 35 bp) 46 animals and homozygote genotype A2A2 (86 bp, 35 bp) 16 animals.



Picture 1: Representatively results of analysis PCR-RFLP for *CSN2* gene by *DdeI* on 3 % agarose gel

- 1, 6 - marker 50 bp DNA Ladder (Fermentas)
- 2 - genotype A1A1(121bp,)
- 3,5 - genotype A2A2 (86 bp, 35 bp)
- 4 - genotype A1A2 (121 bp, 86 bp, 35 bp)
- 7 - PCR product (121 bp)



Scheme 1: Schematic image of PCR-RFLP product *CSN2* gene

- 1 - DNA ladder 50 bp,
- 2 - genotype A1A1 (121 bp)
- 3 - genotype A1A2 (121 bp, 86 bp, 35 bp)
- 4 - genotype A2A2 (86 bp, 35 bp)

Table 1.

Frequency of genotypes and alleles of *CSN2* gene in the population of cattle

Frequency	Genotypes (n =89)			Allels		χ^2 d.f. = 2	p
	A1A1	A1A2	A2A2	A1	A2		
Absolute	27	46	16	100	78	0.2243	0.8939
Relative	0.3034	0.5168	0.1798	0.5618	0.4382		

Frequencies of genotypes and alleles determined in the total population (89 animals), are presented in Table 1. In the total population of cattle heterozygotes A1A2– 0.5168 were the most frequent, while homozygotes A2A2 – 0.1798 were the least frequent ones. This suggests a slight superiority of allele A1– 0.5618.

Genetic equilibrium of analysed population was evaluated on the base χ^2 -test. In the population included in the study non-significant differences in frequencies of genotypes were found. Frequencies of alleles in our population were similar.

Frequencies of A1 allele in our population were similar to those of *CSN2* gene as reported by Bech et al. (1990) for Black-and-White breed and Ehrmann (1997) for Red-and-White breed. Ikonen (1997) reported slight superiority of allele A1 for Ayrshire breed.

The higher frequency of the allele A2 in population of Pinzgau cattle was reported by Beja-Pereira et al. (2003).

Manga et al., (2006) present lower frequency of the allele A1 in population of Czech Spotted and Czech Holstein breed.

Conclusions

It may be concluded that CSN2 is a polymorphic gene with a slight superiority of genotype A1A2 and a slight superiority of allele A1. Genetic structure examined in population of Slovak Pinzgau cattle remained within the range quoted in literature for other cattle breeds.

Sources of research financing: project VEGA no 1/4440/07

References

1. Aschaffenburg, R., 1961, Inherited casein variants in cow's milk, *Nature*, **192**, 431-432
2. Aschaffenburg, R., Sen, A., Thompson, M.P., 1968, Genetic variants of casein in Indian and African zebu cattle, *Comparative Biochemistry and Physiology*, **25**, 177-184
3. Bech, A.M., Kristiansen, K.R., 1990, Milk protein polymorphism in Danish dairy cattle and the influence of genetic variants on milk yield, *J Dairy Res*, **57**, 53-62
4. Beja-Pereira, A., Luikart, G., England, P.R., Bradley, D.G., Jann, O.C., Bertorelle, G., Chamberlain, A.T., Nunes, T.P., Metodiev, S., Ferrand, N., Erhardt, G., 2003, Gene-culture coevolution between cattle milk protein genes and human lactase genes, *Nature Genetics*, **35**, 311 – 313
5. Chin, D., Ng-Kwai-Hang, K.F., 1997, Application of mass spectrometry for the identification of genetic variants of milk proteins, *Milk Protein Polymorphism, IDF, International Dairy Federation, Brussels, Belgium, Special Issue*, **9702**, 334-339
6. Damiani, G., Pilla, F., Leone, P., Caccio, S., 1992, Direct sequencing and bidirectional allele specific polymerase chain reaction of the bovine β -casein B variant, *Animal Genetics*, **23**, 561–566
7. Ehrmann, S., Bartenschlager, H., Geldermann, H., 1997, Quantification of gene effects on single milk proteins in selected groups of dairy cows, *J Anim Breed*
8. *Genet*, **114**, 121–132
9. Elliott, R.B., Reddy, S.N., Bibby, N.J., Kida, K., 1988, Dietary prevention of diabetes in the non-obese diabetic mouse, *Diabetologia*, **31**, 62–64
10. Farrell, H.M. Jr., Jimenez-Florez, R., Bleck, G.T., Brown, E.M., Butler, J.E., Creamer, L.K., Hicks, C.L., Hollar, C.M., Ng-Kwai-Hang, K.F., Swaisgood, H.E., 2004, Nomenclature of the proteins of cows' milk – sixth revision, *Journal of Dairy Sciences*, **87**, 1641–1674
11. Ferretti, L., Leone, P., Sgaramella, U., 1990, Long range restriction analysis of the bovine casein gene, *Nuc. Acids Res.*, **18**, 6829-6833
12. Han, S. K., Shin, Y. C., Byun, H. D., 2000, Biochemical, molecular and physiological characterization of a new β -casein variant detected in Korean cattle, *Anim. Gent.*, **31**, 49–51
13. Ikonen, T., Ojala, M., Ruottinen, O., 1997, Effects of beta and kappa-casein

genotypes on first lactation milk production traits in Finnish Ayrshire cows, *Milk protein polymorphism. Int Dairy Fed*, 47–53

14. Jann, O., Ceriotti, G., Caroli, A., Erhardt, G., 2002, A new variant in exon VII of the bovine β -casein gene (CSN2) and its distribution among European cattle breeds, *J. Anim. Breed. Genet.*, **119**, 65–68

15. Kamiński, S., Cieślińska, A., Kostyra, E., 2007, Polymorphism of bovine beta-casein and its potential effect on human health, *J Appl Genet*, **48**, 189–198

16. Keating, A.F., Smith, T.J., Ross, R.P., Cairns, M.T., 2008, A note on the evaluation of a beta-casein variant in bovine breeds by allele-specific PCR and relevance to β -casomorphin, *Irish Journal of Agricultural and Food Research*, **47**, 99–104

17. Kiddy, C.A., Peterson, R.F., Kopfler, F.C., 1966, Genetic control of the variants of β -casein A, *J. Dairy Sci.*, **49**, 742 (abstr.).

18. Lien, S., Alestrom, P., Klungland, H., Rogne, S., 1992, Detection of multiple β -casein alleles by amplification created restriction sites (ACRS), *Animal Genetics*, **23**, 333–338

19. Manga, I., Řiha, J., Dvořák, J., 2006, Porovnanie vplyvov znakov CSN3 a CSN2 na mliekovú úžitkovosť českého strakatého a holštajnského dobytku testované pri prvej, piatej a ďalších laktáciách, *Acta fytotechnica et zootechnica*, **Mimoriadne číslo**, 13-15

20. McLachlan, C.N., 2006, Breeding and milking cows for milk free of β -casein A1, *United States Patent 7094949*

21. Panicke, L., Freyer, G., Erhardt, G., 1997, Effects of milk protein genotypes on milk production traits, *48th Annual Meeting of the European Association for Animal Production am 25.-28.08.1997 in Vienna/Austria*.

22. Ribadeau-Dumas, B., Brignon, G., Grosclaude, F., Mercier, J.C., 1972, Structure primaire de la caséine β bovine. Séquence complète, *European Journal of Biochemistry*, **25**, 505-514

23. Sambrook, J., Fritsch, E. F., Maniatis, T., 1989, *Molecular cloning: A laboratory manual*. 2nd ed., Cold Spring Harb. Lab. Press, USA. 1989.

24. Stewart, A.F., Bonsing, J., Beattie, C.W., Shah, F., Willis, I.M., Mackinley, A.G., 1987, Complete nucleotide sequences of bovine α s2- and β -casein cDNAs: comparison with related sequences in other species, *Molecular Biology and Evolution*, **4**, 231–241

25. Visser, S., Slangen, C. J., Lagerwerf, F. M., Vandongen, W. D., Haverkamp, J., 1995, Identification of a new genetic variant of bovine β -casein using reversed-phase high-performance liquid chromatography and mass spectrometric analysis, *J. Chromat. A.*, **711**, 141–150

26. Visser, S., Slangen, C.J., Rollema, H.S., 1991, Phenotyping of bovine milk proteins by reversed-phase high performance liquid chromatography, *Journal of Chromatography*, **548**, 361-370

27. Voglino, G.F., 1972, A new β -casein variant in Piedmont cattle, *Animal Blood Groups and Biochemical Genetics*, **3**, 61-62